Storage of Cottonseed and Peanuts Under Conditions Which Minimize Spectrophotometric Changes in the Extracted Oil

WALTER A. PONS, JR., MILDRED D. MURRAY, ROBERT T. O'CONNOR, and JOHN D. GUTHRIE, Southern Regional Research Laboratory,¹ New Orleans, Louisiana

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I N a previous paper (1) it was shown that cotton-seed and peanuts could be stored without change in the free fatty acid content of the oil by adjusting the moisture content to 8% or less and placing the seed in a closed container at 1° or -18° C. In the past the development of free fatty acids and loss of germination have been almost the only indexes used for following the deterioration of seeds in storage. The work to be reported has been directed toward the establishment of other indexes of storage deterioration and toward the further verification of previous recommendations (1) for the storage of experimental samples of cottonseed and peanuts.

Since it seemed likely that the development of the red gossypol of Podolskaja (2) would serve as a useful index of storage changes in cottonseed, this was investigated. Podolskaja has shown that this red pigment increases during the storage of cottonseed and the present work demonstrates its usefulness in cottonseed storage investigations. According to Boatner (6), the red gossypol preparations of Podolskaja were mixtures of gossypol and a purple pigment which Boatner purified and named gossypurpurin. The term red gossypol is used in this paper as a convenient name for a substance or substances which exhibit absorption maxima in chloroform at 528 and 565 m μ , with no intention of implying a relationship to gossypol.

It was also found that an increase in the absorption of the extracted oil in the region of diene conjugation, 236 m μ , is another index of storage change. Since it was found that gossypol has its strongest absorption at 236 m μ , it was necessary to determine the spectrophotometric characteristics of this cottonseed pigment so that the nongossypol absorption at 236 mu could be calculated. This increased absorption at about 236 $m\mu$ is not limited to cottonseed since the extracted oil from peanuts stored at room temperature for over four years also showed a much higher absorption in this region than that from peanuts stored for the same period at 1° or -18° C. The increase in this nongossypol absorption at 236 m μ is, most likely, a result of the development of diene conjugated systems within the unsaturated fatty acid molecules of the oil, by isomerization of the linoleic acid or by peroxidation and probably accompanying dehydration of the oleic acid.

Description of Cottonseed and Peanuts

The varieties of cottonseed used and the moisture contents at the time of storage are given in Table I. Lots 1 to 10, from the U. S. Cotton Field Station at Greenville, Texas, were from cotton picked the day the boll opened. The seed cotton was dried indoors before ginning. Lots 1 to 5 were from early opening bolls and lots 6 to 10 from late opening bolls collected 10 days later. Lots 11 to 16, cottonseed grown at Stoneville, Mississippi, were from the 1945 crop and were shipped to this laboratory as soon as possible after picking.

Lots 17 and 18 were peanuts in the shell from the Georgia Agricultural Experiment Station. The varieties were Tennessee Red and Spanish, respectively. Some storage experiments on these lots of peanuts have been reported previously (1).

TABLE I **Variety** of Cottonseed and 5Ioisture Content of Seed at Time of Storage

Lot No.	Variety	Moisture		
		%		
1	Deltapine 14-060	9.4		
	Station C-42	9.5		
	Acala $W.29.1$	8.9		
	AOL 16-5-8	9.0		
	Mebane Buckellew	9.1		
	Deltapine 14-060	4.6		
	Station C-42	4.3		
	Acala $W-29-1$	5.1		
	AOL 16-5-8	4.8		
	Mebane Buckellew	5.0		
11	Bobshaw 1	7.0		
	Coker 100-9	6.6		
	Delfos 531 C	6.4		
	Deltapine 14-060	7.1		
	Stoneville 2B	6.8		
	Wilds 17	5.5		

Methods

1111 order to investigate the development of red gossypol during storage a chloroform extraction was used since red gossypol is relatively stable in this solvent, tIowever, chloroform extracts could not be used for measurements in the far ultraviolet where chloroform absorbs, and optically pure cyclohexane extracts were used for measurements in this region. In some preliminary work Soxhlet extractions were made and the extracted oil dissolved in cyclohexane prior to spectrophotometric measurements.

Chloroform extraction. The cottonseed was cracked in a Bauer mill without drying or preheating and was screened to separate the meats. The meats were ground in a small Wiley mill using a 20-mesh screen. Immediately after grinding, a 2-g. or 5-g. sample of the ground meats was placed in a Waring Blendor with 30 ml. of chloroform and blended for 15 minutes. The material adhering to the sides was washed down three times with more chloroform during the extraction. The slurry was washed into a 100-ml. wide-mouth volumetric flask, adjusted to room temperature in a beaker of water, made to volume with chloroform, and filtered through a coarse filter paper, rejecting the first portion of the filtrate. The cloudy filtrate was refilfered. A portion of the filtrate, 20 nil., was pipetted into a tared flask, the chloroform evaporated on a steam bath, the weight of the oil determined and calculated to oil concentration in grams per liter. Spectrophotometric determinations of the optical densities at various wave lengths were made on the extracts with a Beckman speetrophotometer using a 1-cm. cell. The data were calculated to

¹ One of the laboratories of the Bureau of Agricultural and Indus
trial Chemistry, Agricultural Research Administration, U. S. Depart
ment of Agriculture.

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extinction coefficients based on the oil concentration of the extracts in grams per liter and a 1-era. cell length. The extractions and speetrophotometrie measurements were made on a strict time schedule.

Cyclohexane extraction. This procedure was identical with the above chloroform extraction except that optically pure cyelohexane replaced the chloroform.

Soxhlet extraction with ethyl ether or with lowboiling petroleum ether. The cottonseed meats or peanut kernels were ground in a food cutter provided with a 19-tooth blade. Immediately after grinding, a 10-g. sample was weighed into a paper thimble and extracted in a medium-sized Soxhlet extractor for four hours with peroxide-free ethyl ether or with low-boiling petroleum ether (A.O.C.S. Specification H-2-41). The material was removed from the thimble and ground to a fine powder in a mortar. It was then returned to the original thimble and the extraction continued for an additional 16 hours. Most of the solvent was evaporated on a steam bath and the last traces were removed in a vacuum desiccator. A]-g. sample of the extracted oil was weighed into a 25-ml. beaker, dissolved in optically pure cyclohexane, filtered into a 25-ml. volumetric flask, the paper was washed with eyclohexane, and the filtrate made to volume. This solution, which had a concentration of 40 g. of oil per liter, was used for speetrophotometric measurements in the near ultraviolet and in the visible regions. A 0.1-g. sample of the oil was treated in the same way to give a solution having a concentration of 4 g. per liter. Portions of this were diluted quantitatively to give solutions having concentrations of 0.4 g. per liter and 0.04 g. per liter. These were used for spectrophotometric measurements in the far ultraviolet.

Preparation of Pure Gossypol

In order to calculate the nongossypol absorption of the cyclohexane extracts of cottonseed, it was necessary to know the speetrophotometrie characteristics of pure gossypol in eyclohexane. Pure gossypol was prepared by the following procedure from cottonseed pigment glands separated by the flotation process $(3, 4).$

Cottonseed pigment glands, 250 g., were extracted with 1 liter of acetone for three hours on a mechanical shaker. After filtration through a hardened paper, the volume of the filtrate was reduced to 600 ml. under reduced pressure. The filtrate was mixed with 600 ml. of glacial acetic acid and the mixture allowed to stand overnight. The precipitate was separated by filtration, washed by suspension in low-boiling petroleum ether, and again separated by filtration. Its weight was 101 g. This crude gossypol acetic acid was dissolved in 800 nil. of acetone, filtered, and 400 ml. of glacial acetic acid was added with stirring. After 2 hours the precipitate was separated by filtration, washed by suspension in low-boiling petroleum ether, and refiltered. The precipitate was again dissolved in acetone and again precipitated with acetic acid in the same way. The weight of the precipitate was 91 g.

The gossypol acetic acid was converted to free gossypol by suspending it in 600 ml. of peroxide-free ethyl ether and pouring it into a large evaporating dish containing 500 ml. of water and a small amount of sodium hydrosulfite (5). The ether was renmved on the steam bath, while stirring with a thermometer, care being taken that the temperature of the water did not exceed 50° C. After the ether was removed, the lumps of free gossypol were crushed with a pestle, transferred to a Biichner funnel, and washed with water. The weight of material was 87 g.

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ซ COEFF 100 ZOLUZ EXT **50** - **r~l I I I** 0 **200 300 400 500** WAVELENGTH MU

FIG. 1. Ultraviolet absorption spectrum of gossypol in cyclohexane.

The crude gossypol was dissolved in 800 ml. of peroxide-free ether, an equal volume of low-boiling petroleum ether was added, and the solution concentrated under reduced pressure until crystallization began. One-half volume of low-boiling petroleum ether was then added, and after one hour the precipitate was separated by filtration. A second crop of material was obtained by evaporation to 500 ml. under reduced pressure. This second crop was combined with the first, dissolved in ether, and precipitation by the addition of low-boiling petroleum ether was carried out two more times. The weight of material was now 58 g.

To effect further purification, the material was dissolved in ethyl ether, converted to gossypol acetic acid, brought back to free gossypol, and recrystallized three times from a mixture of ethyl ether and petroleum ether in a manner similar to that given above. It was then dissolved in acetone and precipitated as gossypol acetic acid. The gossypol acetic acid was dissolved in ethyl ether and converted to free gossypol in the manner described. The pure gossypol was dried in a vacuum oven at 50° C. for four hours and placed overnight in a vacuum desiccator over phosphorus pentoxide. The yield was 33 g.

The analysis (micro) of this material gave the following values :

Carbon 69.0, hydrogen 5.9. Calculated for $C_{30}H_{30}O_8$: carbon 69.5, hydrogen 5.8. The nitrogen content by Kjeldahl digestion and Nesslerization was 0.05% .

A speetrophotometric curve made on this pure gossypol in cyelohexane is shown in Fig. 1. Spectrophotometric data to establish the extinction coefficients 2 at the three maxima are given in Table II. For comparison, data on a preparation made by the method of F. H. Smith (5), on a preparation supplied by F. H. Smith, and on two preparations supplied by Roger Adams, are given. The close agreement of the **spectrophotometric data for all these preparations is good evidence for the purity of all of them. The agreement in absorption characteristics of freshly prepared and old samples is evidence that gossypol is reasonably stable when protected from light under ordinary laboratory conditions.**

Calculation of Nongossypol Absorption at 236 m_µ

If we assume that practically all of the absorption of the extracts of eottonseed in cyclohexane at 358 mu or at 286 mu is due to gossypol, it is possible to calculate the absorption due to gossypol at $236 \text{ m}\mu$ **by use of the appropriate factors and obtain the non**gossypol absorption at 236 m_{μ} by difference. Al**though it is realized that the values so obtained arc somewhat empirical, their relative values are valid and serve as a good index of change in storage. The data reported in Table II for the absorption of gossypol in cyclohexane show that the ratio of extinction** at 236 m μ to that at 286 m μ is 2.96. However, 2.9 **has been used in the calculations in order to avoid negative values for nongossypol absorption. It is likely that a very small part of the absorption at** $286 \text{ m}\mu$ is due to substances other than gossypol; so the calculated gossypol absorption at $236 \text{ m}\mu$ based on the value at $286 \text{ m}\mu$ is likely to be higher than **the true absorption. The ratio of extinction at 236** m_{μ} to that at 358 m_{μ} averages 5.19; so for purposes **of calculation the value 5.2 has been used to obtain** the gossypol absorption at 236 $m\mu$ from the absorption at 358 m_p. Thus two formulas have been used to calculate the nongossypol absorption at $236 \text{ m}\mu$.

1. Nongossypol absorption at 236 $m\mu = a_{236}-2.9 a_{256}$

2. Nongossypol absorption at 236 $m_{\mu} = a_{236} - 5.2 a_{358}$ **a=extinction coefficient (concentration in g. per liter, solution depth 1 cm.).**

2All quantitative values of absorption referred to in this paper are given as extinction coefficient (alpha) defined as $\text{Log } \frac{10}{1}$ /cl. numerically

equal to the expression $E_{1 \text{ cm}}^{g/1}$ **, where I_o and I are the intensities of the incident and transmitted light respectively, c is the concentration in grams per liter, and 1 is the cell length in centimeters.**

Results with Cottonseed

Considerable preliminary work, not to be reported in detail, was done by Soxhlet extraction with peroxide-free ethyl ether or low-boiling petroleum ether. The nongossypol absorption values at $236 \text{ m}\mu$ obtained **by these methods indicated that storage at low temperatures retarded or prevented the increase in nongossypol absorption. Itowever, the values were** quite **variable and the data contained** a number **of inconsistencies. Therefore, only values for nongossypol absorption at 236 mg obtained by the cyclohexane method are given siuee this method gives more reproducible and consistent results. However, some of the ethyl ether extraction values are of interest in relation to the gossypol content of the seed since this method has been used for the quantitative extraction of gossypol. Table III shows the absorption values**

TABLE III	

Spectrophotometrie Characteristics of Ethyl Ether Extracts of Cottonseed Stored at Two Different Temperatures

at $358 \text{ m}\mu$, one of the maxima of gossypol. These **iudicate that little or no change in gossypol content of the cottonseed took place during storage for 14** months at 27° or at 1°C. Table III also shows that **an iucrease in absorption of the ethyl ether extracted** oil at 560 μ occurred in lots 1 to 5 stored at 27 $^{\circ}$ C. **These lots, from the early opening bolls, had a mois**ture content of about 9% when stored. This increase in absorption at 560 $m\mu$ is probably related to the **increase in red gossypol, which is better demonstrated by chloroform extraction. This increase in absorption** at 560 m μ did not occur in seeds stored at 1^o C., nor **did it occur in lots 6 to 10, the late opening bolls of the series, which had a moisture content of about 5% when stored.**

The spectrophotometric characteristics of the chloroform extracts for lots 1 to 10 are shown in Table

IV. A definite increase in red gossypol, as indicated by absorption at 565 m μ or at 528 m μ , took place at 27° C. This increase occurred in all 10 lots of seed at 27° C. but was much larger in the seed from the early opening bolls, which had a rather high moisture content. No significant change in the red gossypol absorption took place in the seed stored at 1° C. The absorption values at 365 m_{μ} do not indicate any very large change in the gossypol content of the seed under either of the storage conditions.

Extensive data were obtained on lots 11 to 16 which were stored at 27° , 1°, and -18° C. and analyzed at intervals of 5, 7, and 18 months. Complete spectrephotometric curves were made on the chloroform extracts of lot 11. Some of these are shown in Fig. 2 to illustrate the large increase in red gossypol, absorption maxima at 565 m μ and 528 m μ , which took place during storage for 18 months at 27° C. Fig. 2 also shows that very little change in red gossypol took place in the samples stored at 1° and at -18° C. This is borne out by the data in Table V. A significant increase in absorption at 565 m μ and at 528 m μ is evident after five months at 27° C. and is quite large after 18 months at this temperature. It is worthy of note that lot 16, which showed the smallest increase in red gossypol at 27° C., had the lowest moisture content at time of storage (Table I). No significant change can be demonstrated at 1° or at -18° C. The data for the absorption of the chloroform extracts at 365 m_{μ} are indicative of little or no change in the amount of gossypol.

Table VI shows Evelyn colorimeter measurements in the region of red gossypol absorption of independent chloroform extracts of lots 11 to 16. They are entirely in agreement with the spectrophotometric measurements and show that a photoelectric colorimeter can be used to follow the development of red gossypol during storage of cottonseed.

Fig. 3 shows spectrophotometric curves on the cold cyclohexane extract of lot 11 at the time of storage and after 18 months at 1° and at -18° C. An increase in absorption at 236 $m\mu$ is evident after storage at 27~ That this change is real and significant is brought out by Tables VII and VIII. Table VII gives the spectrophotometric absorption characteristics of the cyclohexane extracts of lots 11 to 16. Although an increase in absorption at 236 m μ is evident in storage at 27° C., such a large part of the 236 m_{μ} absorption is due to gossypol, variable amounts of which may be extracted by the cyclohexane, it is best

to calculate the nongossypol absorption by the formulas given earlier in this paper and base the conclusions on these calculations. Table VIII shows the nongossypol absorption values at 236 m μ . A significant increase in the nongossypol absorption at 236 m μ , the region of diene conjugation, is evident after five months at 27° C. This increase is much greater after 18 months. No significant increase in the nongossypol absorption at 236 m μ took place at either 1° or -18° C. However, the average value at 1°C, tends to be a little higher after 18 months than the original value. It would probably be best, therefore, to store cottonseed at -18° C. if the storage period was long and one wished to be sure that no change took place in the nongossypol absorption of the extracted oil at $236 \; \text{m}$ u.

* Oil concentration not obtained due to accident.

Lot No. Orig. [5 Months . 7 :Months 18 **Months** $\texttt{Sample} \begin{array}{|l|} \texttt{27°C,} & -18^{\circ}\text{C.} & 27^{\circ}\text{C.} & 1^{\circ}\text{C.} & -18^{\circ}\text{C.} & -18^{\circ}\text{C.} & 27^{\circ}\text{C.} & 1^{\circ}\text{C.} & -18^{\circ}\text{C.} \end{array}$ $Alpha~565$ m $µ$ 11 ... 12 ... 13 ... 14 ... 15 ... 16 ... **Average** 0.008 0.011 0.007 0.009 0.009 0.005 0.008 0,012 0.013 0.011 0.012 0.014 0.008 0.012 0,009 0.012 0.010 0.010 0.011 0.005 0.010 0.013 0.(}13 0.012 0.010 0.015 0.005 0.011 0.009 0.010 0.009 0.008 0,010 0.005 0.009 0.010 b.610 0.008 0.008 0.010 0.004 0,008 0.025 0.018 0.018 0,021 0.025 0.008 0.019 0.009 **0.011** 0.009 0,009 0.011 0.005 0.009 0.009 0.010 0.008
0.009
0.010 0.005 0.009 Alpha 528 m μ 11 *...* 12 ... 13 ... 14 ... 15 ... 16 ... **Average** 0.007 0.010 0,007 0,009 0,008 0.005 0.008 0.011 0.012 0.010 0.010 0.012 0.008 0.011 0.008 0.010 0.009 0.009 0.009 0.O05 0.008 $\begin{array}{c} 0.012\ 0.012\ 0.010\ 0.009\ \end{array}$ o,012 0,005 $0,010$ 0.008 0,009 0.009 **o.O(Ig** n.oog o.005 0.008 0.009 0.009
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0.007
0.008 o.009 0.005 0.008 Alpha $365 \text{ m}\mu$ 11 12 13 14 15 1,6 .. Average 1.05 1.01 0.71 1.11 0.90 0.59 0.90 1.16 1.08 0.78 1.08 1.06 1.04 **1.03** $\begin{array}{c} 1.22\ 1.18\ 0.88 \end{array}$ **1.02** 1.04 0.67 **1.00** 1,13 **1.06** 0.88 1.04 1.04 0.65 0.97 1.13 **1.12** 0.79 1.06 0.99 0.70 (}.97 **1.08** 1.11 0,87 1.62 0,97 0.61 0.94 1.21 1.16 0.84 1.11 1,04 0.79 1.03 **1.20** $_{\rm 0.90}^{1.21}$ $_{\rm 1.03}^{\rm 1.09}$ 0.78 1.04 **1.25** 1.16 0.88 $\begin{smallmatrix} 1.10 0.98 \end{smallmatrix}$ 0.85 1.04

TABLE V **Spectrophotometric Characteristics of Chloroform Extracts of Cottonseed Stored for Various Times at Different Temperatures.**

Results with Peanuts

In order to see whether the increase in absorption at about 236 m_{μ} took place in some oil seed other **than cottonseed, two lots of peanuts that had been in storage over four years were investigated. It was** found that the maximum for extinction in this region was 227 to 234 $m\mu$ in the peanut extracts. The inter**pretation of the data was much simpler than with cottonseed since gossypol was absent. The data in Table IX show that the absorption in the region of diene conjugation is much higher in the oil extracted** from the peanuts stored at 27°C, than from those stored at 1° or -18° C. Unfortunately, no original **value was available for these samples.**

Preliminary experiments on the stability of the oil from these peanuts indicated that the oil from peanuts stored at 27°C. for over 4 years was much less **stable than that from peanuts stored at the lower**

temperatm'es. The oils all had low peroxide values at the start of the stability experiments.

TABLE VI Evelyn (?olorimeter **Measurements of ('hh)roform Extracts** ef **Cottonseed Stored at l)iffereot Temperatures**

	"E" Values with 565 Filter							
Lot.	Orig.	7 Months		18 Months				
No.		27° C.	1° C.	-18° C.	27° C.	1° C.	-18° C.	
11 12	0.010 0.013	0.014 0.015	0.010 0.012	0.011 0.012	0.030 0.023	0.014 0.014	0.011 0.014	
13 14	0.009 0.010	0.013 0.014	0.010 0.011	0.009 0.009	0.019 0.025	0.011 0.011	0.010 0.010	
15 16	0.011 0.005	0.016 0.007	0.012 0.005	0.012 0.006	0.026 0.009	0.012 0.006	0.012 0.006	
Av.	0.010	0.013	0.010	0.010	0.022	0.011	0.011	

The peanuts stored at 27°C. for over four years were not viable while those stored at 1° and -18° C. germinated 100% . The testa of the kernels of the

Spectrophotometric Characteristics of Cyclohexane Extracts of Cottonseed Stored for Various Times at Different Temperatures.

*** These values obtained after eight months, since the values at seven months were in error,**

TABLE Vlll

* These values **obtained after eight months, since the values at seven months were in** error.

peanuts stored at 27^oC. were much darker than the **testa of those stored at the lower temperatures.**

Acknowledgments

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Summary

1. Cottonseed was stored for over a year without appreciable change in red gossypol content by ad**justing the moisture content to 8% or less and plac**ing in closed containers at 1° or at -18° C. Cottonseed stored at room temperature, 27°C., showed a **large increase in red gossypol content.**

2. Cottonseed stored in the above manner at 1^o or at -18°C, showed little or no increase in the nongossypol absorption of the extracted oil at 236 m_{μ} , **the region of diene conjugation. This property of the** extracted oil showed a large increase when cottonseed was stored at room temperature, 27°C.

3. Gossypol dissolved in cyclohexane has spectrophotometric extinction coefficients (concentration in g. per

Fro. 3. **Absorption spectra of eyclohexane extracts** of **lot 11 cottonseed.**

Curve 1. Original seed.

Curve 2. After storage for 18 months at 1° and -18° C.

Curve 3. After storage for 18 months at 27^cC.

liter, solution depth 1 era.) of 207, 70, and 40 at the maxima at 236 m μ , 286 m μ , and 358 m μ , respectively.

4. The extracted oil from peanuts stored for over four years at room temperature, 27°C., showed a much greater absorption in the region of $227 \text{ m}\mu$ to 234 m μ than did peanuts stored for the same time at 1° or at -18° C.

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